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THE FIRST CASE OF TRIPLOIDY IN SKINKS OF THE GENUS *Ophiomorus* (REPTILIA: SCINIDAE)

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Polyploidy is a well-known phenomenon among various groups of animals, plants and fungi. However, the occurrence and study of this phenomenon among reptiles is still underestimated. Currently, polyploid (3 - 4n) and mosaic (2/3n) individuals are known in eleven families of reptiles. The paper presents a unique case of polyploidy in a skink of the genus *Ophiomorus*. Using DNA flow cytometry, we detected diploid and triploid specimens of *O. tridactylus* in a population near Jask in Southern Iran. This may be the result of hybridization or natural spontaneous autotriploidy. Further studies of these lizards in the region will make it possible to determine the causes of this phenomenon.

Keywords: polyploidy; DNA flow cytometry; genome size; lizards; Iran.

INTRODUCTION

Polyploidy is interesting, but poorly studied phenomena occurring in some animal, plant, and fungal lineages (Mason and Pires, 2015; Dar and Rehman, 2017). Two categories of polyploids are able to be recognized: allopolyploids are hybrids and autopolyploids have chromosome sets derived from a single species (Borkin et al., 1996). Polyploids have been documented across a wide range of vertebrate species (Gregory and Mable, 2005). In reptiles, polyploidy is relatively common. Triploid species and lineages have been identified in about twenty species complexes of seven families: Agamidae Spix,

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1825, Gekkonidae Gray, 1825, Gymnopthalmidae Fitzinger, 1826, Liolaemidae Frost et Etheridge, 1989, Scincidae Gray, 1825, Teiidae Gray, 1827, and Typhlopidae Merrem, 1820 (Bogart, 1980; Kearney et al., 2009; Trifonov et al., 2015; Abdala et al., 2016; Stöck et al., 2021). Their origin is usually associated with interspecific hybridization and parthenogenetic reproduction (Kearney et al., 2009). Sometimes, triploid parthenogenetic females interbreed with males of diploid bisexual species, which can result in polyploid individuals. Such cases were noted in the genus Aspidoscelis Fitzinger, 1843 (Teiidae), where hybridization of triploid parthenogenetic females with diploid males can lead to the formation of tetraploid offspring (Hardy and Cole, 1998; Lutes et al., 2011; Cole et al., 2014, 2017). Sometimes, in the genus Darevskia Arribas, 1999 (Lacertidae Bonaparte, 1831) diploid parthenogenetic females interbreed with males of bisexual species, which can lead to the formation of triploid and tetraploid hybrids (Darevsky et al., 1973; Danielyan et al., 2008; Freitas et al., 2019).

Occasionally, species with sharp differences in the number of chromosomes can be observed within a reptilian family. However, such differences are not always associated with polyploidy. Apparently, a similar case was observed in the family Chamaeleonidae Rafinesque,

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1815, where *Rieppeleon brevicaudatus* (Matschie, 1892) has 62 chromosomes, while rest species of the family have 28 – 38 chromosomes (Mezzasalma et al., 2023). Some authors (Hall, 1970) believed that this case could be associated with polyploidy, but recently such an increase in the number of chromosomes is associated with massive chromosomal rearrangements (Mezzasalma et al., 2024).

Non-hybrid mixoploidy (2/3n) and spontaneous autopolyploidy (3n) were also observed among reptilian species. A sex-related diploid-triploid mosaicism was observed in populations of two bisexual species: Platemys platycephala (Schneider, 1792) from the family Chelidae Gray, 1831 and Liolaemus chiliensis (Lesson, 1830) from the family Liolaemidae. In both species, triploidy was positively or negatively (respectively) associated with the number of males and could therefore play a role in sex determination (Bickham et al., 1985; Lamborot et al., 2006; Bickham and Hanks, 2009). Also, a mixture of diploid and triploid somatic cells was observed in a parthenogenetic female of Lepidophyma flavimaculatum Duméril, 1851 from the family Xantusiidae Baird, 1858 (Bezy, 1972). A case of spontaneous autotriploidy has recently been found in Saltuarius cornutus (Ogilby, 1892) from the family Carphodactylidae Kluge, 1967 (Pensabene et al., 2024).

Our study focuses on members of the genus *Ophiomorus* A. M. C. Duméril et Bibron, 1839, which includes 12 species of relatively small skink lizards with short limbs and often reduced fingers and toes (Nabizadeh et al., 2023; Uetz, 2024). The distribution of the genus is disjunct and divided into isolated areas in the Balkans, Turkey, the Levant, Iran, Pakistan and India. 7 - 8 species inhabit Iran, where they live mostly in sand dunes (Šmíd et al., 2014). The karyotype, phylogeny, range, and ecology of these skinks are extremely insufficiently studied. The purpose of this paper is to describe a unique case of triploidy in these rare and poorly studied skinks.

MATERIAL AND METHODS

During a travel to Iran in 2018, we collected twelve adults of O. tridactylus (Blyth, 1853), O. maranjabensis Azemi, Farhadi Qomi, Kami et Anderson, 2011, and O. streeti Anderson et Leviton, 1966 from four localities (Table 1; Fig. 1). The amount of DNA per nucleus (genome size) was measured by DNA flow cytometry. Flow cytometry was performed by using of microscope-based flow fluorimeter with mercury arc lamp as a light source constructed at the Institute of Cytology, Russian Academy of Sciences, St. Petersburg (https://patents.google. com/patent/SU1056008A1/ru). DNA-histograms were acquired with a multichannel analyzer connected with a microcomputer. The analysis rate was 100 - 200 cells per second; four runs were done on each sample, with the total number of cells measured per sample being above 10,000. The peaks of DNA histograms were plotted to the Gaussian curves by means of the least-square technique.

Red blood cells were taken by a micropipette from the heart. Each individual was anaesthetized by chloroform. Tested cells were mixed with reference cells and assayed simultaneously. Spleenocytes of Mus musculus Linnaeus, 1758 (C57B1) males were used as a reference standard. In arbitrary units, genome size of an individual under the study was determined as the ratio of its cell peak mean to that of M. musculus (6.8 pg by Bianchi et al., 1983). The cell samples were suspended in Versene solution (pH 7.3 - 7.7; Biolot, St. Petersburg), with a total cell concentration of approximately 106 cells/ml and stored before study at 4°C. The cells were lysed by addition of Triton X-100 (Ferak, Berlin) at a final concentration of 0.1%, and stained with a mixture of olivomycin (OM, Moscow Medicine Plant) and ethidium bromide (EB, Calbiochem) at the following final concentration: 40 µg/ml OM, 20 µg/ml EB, and 15 mM MgCl₂. The stained samples were measured after 4-6 h. Other details of the methodology were published earlier (Vinogradov et al., 1990; Rozanov and Vinogradov, 1998).

TABLE 1. The Nuclear DNA Content (pg) in Erythrocytes of Three Species of the Genus Ophiomorus

No.	Species	Ploidy	Locality	Coordinate	Altitude -	Nuclear DNA content		
						п	$\text{mean}\pm\text{SD}$	range
1	maranjabensis	2 <i>n</i>	"Safari sands"	34°18' N 51°51' E	827	1	4.564	
2	_''_	2 <i>n</i>	Kashsan	33°53' N 51°49' E	957	1	4.572	
3	streeti	2 <i>n</i>	Kerman	27°56' N 58°05' E	451	5	4.494 ± 0.014	4.468 - 4.502
4	tridactylus	2 <i>n</i>	Bahu Kalat	25°46' N 61°25' E	38	3	4.509 ± 0.010	4.503 - 4.520
5	_''_	2 <i>n</i>	Jask	25°48' N 57°49' E	75	1	4.532	
	''	3 <i>n</i>	_''_	_''_	_''_	1	6.850	

RESULTS AND DISCUSSION

Among two individuals of *O. tridactylus* from Jask, one was diploid and the other was triploid. The nuclear DNA content of the triploid individual was equal to 6.850 pg, while in diploid *O. tridactylus* was 4.532 pg. Genome size in the other *O. tridactylus* population we studied and in two other *Ophiomorus* species ranged from 4.503 to 4.572 pg (Table 1). The triploid specimen was adult, normally developed and had a normal color pattern.

In the family Scincidae, triploidy is known only in the Menetia grevii species complex consisting of 2-3diploid bisexual species (from south-central Australia) and three triploid hybrid parthenogenetic lineages widespread throughout Australia (Adams et al., 2003). Our record of triploidy in the genus Ophiomorus is the second for the family. Unfortunately, due to small number of individuals studied, it is difficult to classify the type of polyploidy (allo- or auto-) which we discovered. It could be the result of hybridization with closely related species (perhaps, O. brevipes; see Fig. 1) or a case of spontaneous autotriploidy (e.g., as a result of the fusion of the first polar nucleus with the nucleus of the secondary oocyte; Stenberg and Saura, 2013). However, among terrestrial vertebrates the latter type of polyploidy is much more common in amphibians than in reptiles (Litvinchuk et al., 2016; Pensabene et al., 2024). Only new studies of the species in the region will help to resolve this issue.

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REFERENCES

Abdala C. S., Baldo D., Juárez R. A., and Espinoza R. E. (2016), "The first parthenogenetic pleurodont iguanian: a new all-female *Liolaemus* (Squamata: Liolaemidae) from Western Argentina," *Copeia*, **104**(2), 487 – 497.



Fig. 1. Map showing ranges of seven species and studied localities of *Ophiomorus* in Iran. The red asterisk marks the location (Jask), where the triploid individual of *O. tridactylus* was found. Violet circles are localities of *O. maranjabensis*, blue circle is the locality of *O. streeti*, and red circle and asterisk are localities of *O. tridactylus*. Numbers for localities are given in Table 1. Photo credit: DAM (*O. tridactylus* from Bahu Kalat).

- Adams M., Foster R., Hutchinson M. N., Hutchinson R. G., and Donnellan S. C. (2003), "The Australian scincid lizard *Menetia greyii*: A new instance of widespread vertebrate parthenogenesis," *Evolution*, 57(11), 2619 – 2627.
- Bezy R. L. (1972), "Karyotypic variation and evolution of the lizards in the family Xantusiidae," *Contrib. Sci.*, 227, 1 – 29.
- Bianchi N. O., Redi C., Garagna C., Capanna E., and Manfredi-Romanini M. G. (1983), "Evolution of the genome size in *Akodon* (Rodentia, Cricetidae)," *J. Mol. Evol.*, 19, 362 – 370.
- Bickham J. W. and Hanks B. G. (2009), "Diploid-triploid mosaicism and tissue ploidy diversity within *Platemys platycephala* from Suriname," *Cytogen. Genome Res.*, 127, 280–286.
- Bickham J. W., Tucker P. K., and Legler J. M. (1985), "Diploid-triploid mosaicism: an unusual phenomenon in sidenecked turtles (*Platemys platycephaia*)," *Science*, 227, 1591–1593.
- Bogart J. P. (1980), "Evolutionary implications of polyploidy in amphibians and reptiles," in: W. H. Lewis (ed.), *Polyploidy: Biological Relevance*, Plenum Press, New York and London, pp. 341 – 378.
- Borkin L. J., Litvinchuk S. N., and Rosanov J. M. (1996), "Spontaneous triploidy in the crested newt, *Triturus cristatus* (Salamandridae)," *Russ. J. Herpetol.*, **3**(2), 152 – 156.
- Cole C. J., Taylor H. L., Baumann D. P., and Baumann P. (2014), "Neaves' whiptail lizard: the first known tetraploid parthenogenetic tetrapod (Reptilia: Squamata: Teiidae)," *Breviora*, **539**, 1 19.
- Cole C. J., Taylor H. L., Neaves W. B., Baumann D. P., Newton A., Schnittker R., and Baumann P. (2017), "The second known tetraploid species of parthenogenetic te-

trapod (Reptilia: Squamata: Teiidae): description, reproduction, comparisons with ancestral taxa, and origins of multiple clones," *Bull. Mus. Comp. Zool.*, **161**(8), 285 – 321.

- Danielyan F., Arakelyan M., and Stepanyan I. (2008), "Hybrids of *Darevskia valentini*, *D. armeniaca* and *D. unisexualis* from a sympatric population in Armenia," *Amphibia–Reptilia*, **29**, 487 504.
- Dar T.-UI.-H. and Rehman R.-UI. (eds.) (2017), Polyploidy: Recent Trends and Future Perspectives, Springer.
- Darevsky I. S., Uzell T. M., Kupriyanova L. A., and Danielyan F. D. (1973), "Triploid hybrid males in sympatric populations of some parthenogenetic and bisexual species of rock lizards of the genus *Lacerta*," *Byull. Mosk. Obshch. Estestvoisp.*, 78, 48 – 58. [in Russian].
- Gregory T. R. and Mable B. K. (2005), "Polyploidy in animals," in: T. R. Gregory (ed.), *The Evolution of the Genome*, Elsevier, pp. 427 – 517.
- Freitas S. N., Harris D. J., Sillero N., Arakelyan M., Butlin R. K., and Carretero M. A. (2019), "The role of hybridisation in the origin and evolutionary persistence of vertebrate parthenogens: a case study of *Darevskia* lizards," *Heredity*, 123, 795 – 808.
- Hall W. P. (1970), "Three probable cases of parthenogenesis in lizards (Agamidae, Chamaeleontidae, Gekkonidae)," *Experientia*, 26(11), 1271 – 1273.
- Hardy L. M. and Cole C. J. (1998), "Morphology of a sterile, tetraploid, hybrid whiptail lizard (Squamata: Teiidae: *Cnemidophorus*)," *Am. Mus. Novitates*, **3228**, 1 – 16.
- Kearney M., Fujita M. K., and Ridenour J. (2009), "Lost sex in the reptiles: Constraints and correlations," in: I. Schön et al. (eds.), *Lost Sex*, Springer, pp. 447 – 474.
- Lamborot M. M., Manzur E., and Alvarez-Sarret E. (2006), "Triploidy and mosaicism in *Liolaemus chiliensis* (Sauria: Tropiduridae)," *Genome*, **49**, 445 – 453.
- Litvinchuk S. N., Borkin L. J., Skorinov D. V., Pasynkova R. A., and Rosanov J. M. (2016), "Natural polyploidy in amphibians," *Vestn. SPb Gos. Univ. Biol.*, 3(3), 77 – 86. [in Russian]
- Lutes A. A., Baumann D. P., Neaves W. B., and Baumann P. (2011), "Laboratory synthesis of an independently reproducing vertebrate species," *PNAS*, **108**(24), 9910 – 9915.
- Mason A. S. and Pires J. C. (2015), "Unreduced gametes: meiotic mishap or evolutionary mechanism?" *Trends Genet.*, 31(1), 5 – 10.

- Mezzasalma M., Streicher J. W., Guarino F. M., Jones M. E. H., Loader S. P., Odierna G., and Cooper N. (2023), "Microchromosome fusions underpin convergent evolution of chameleon karyotypes," *Evolution*, 77(9), 1913 – 1944.
- Mezzasalma M., Macirella R., Odierna G., and Brunelli E. (2024), "Karyotype diversification and chromosome rearrangements in squamate reptiles," *Genes*, **15**, 371.
- Nabizadeh H., Rastegar-Pouyani N., Rastegar-Pouyani E., and Rajabzadeh M. (2023), "Interspecific variation within the genus *Ophiomorus* Duméril & Bibron, 1839 (Sauria: Scincidae) in Iran based on morphological characters," *Iran. J. Animal Biosyst.*, **18**(1), 47 – 70.
- Pensabene E., Augstenová B., Kratochvíl L., and Rovatsos M. (2024), "Differentiated sex chromosomes, karyotype evolution, and spontaneous triploidy in carphodactylid geckos," *J. Hered.*, esae010.
- **Rozanov Y. M. and Vinogradov A. E.** (1998), "Precise DNA cytometry: investigation of individual variability in animal genome size," *Tsitologiya*, **40**, 792 800. [in Russian]
- Šmíd J., Moravec J., Kodym P., Kratochvíl L., Yousefkhani S. S. H., Rastegar-Pouyani E., and Frynta D. (2014), "Annotated checklist and distribution of the lizards of Iran," *Zootaxa*, 3855, 1 – 97.
- Stenberg P. and Saura A. (2013), "Meiosis and its deviations in polyploid animals," *Cytogen. Genome Res.*, 140(2-4), 185-203.
- Stöck M., Dedukh D., Reifová R., Lamatsch D. K., Starostová Z., and Janko K. (2021), "Sex chromosomes in meiotic, hemiclonal, clonal and polyploid hybrid vertebrates: along the 'extended speciation continuum'," *Phil. Trans. R. Soc. B*, **376**, 20200103.
- Trifonov V. A., Paoletti A., Caputo Barucchi V., Kalinina T., O'Brien P. C. M., Ferguson-Smith M. A., and Giovannotti M. (2015), "Comparative chromosome painting and NOR distribution suggest a complex hybrid origin of triploid *Lepidodactylus lugubris* (Gekkonidae)," *PLoS ONE*, 10(7), e0132380.
- Uetz P. (ed.) (2024), *The Reptile Database*, http://www.rep-tile-database.org (accessed February 28, 2024).
- Vinogradov A. E., Borkin L. J., Günther R., and Rosanov J. M. (1990), "Genome elimination in diploid and triploid *Rana esculenta* males: cytological evidence from DNA flow cytometry," *Genome*, 33, 619 – 627.