

Cilia-like structure, primary cilium and mechano-transduction in the osteocyte

Authors

Rustem E. UZBEKOV¹, Claude Laurent BENHAMOU²

¹Laboratoire des microscopies—Department of Microscopy, Faculty of Medicine, François Rabelais, University, Tours, 37032 Tours Cedex 1, France

Phone: +33-247-366071 or +33-234-379692, Fax: +33-247-478207, E-mail: rustem.uzbekov@univ-tours.fr

²IPROS, Hôpital Porte Madeleine, 45032 Orléans Cedex 02, France

Phone: +33, E-mail: benhamou.cl@wanadoo.fr

The centrosome represents the real core of microtubule organization in cells. The centrosome is involved in organelles such as cilia and flagella. The osteocyte is the bone cell in charge of mechano-reception and mechano-transduction in the skeleton [1, 2, 3, 4]

Primary cilia are present on most eukaryotic cells, and are considered to play a role in mechanosensing, particularly in the translation from mechano-reception to biochemical signals [5, 6, 7]. This role has been largely studied in the renal tubule epithelium; in hepatocytes, myocytes, red blood cells [6, 8, 9].

The primary cilium is a part of the cytoskeleton, it is characterized by a microtubule organization [10], under dependence of the centrosome. The centrosome is an organelle responsible of the microtubule organizing centers. It is involved in the mitotic spindle generation of the cell division, and in the function of primary cilium.

It is constituted of two centrioles: the mother and the daughter centrioles. Its molecular structure is mainly acetylated tubulin [11] with a cylinder shape of nine triplets in the mother centriole.

Our experiment [12] was developed on male Wistar rats, 26 weeks old, specifically on the upper part of the tibia, to study the osteocytes of cortical bone, with a particular care to the cell orientation. The study involved microscopy immunostaining (acetylated alpha-tubulin coupled to confocal microscopy), and TEM (transmission electron microscopy).

How many osteocytes possess such a structure? From 236 cells we observed, 222 were positively immunostained (94 %) [12]. It remains to be defined if the 6 % without staining of the centrosome could be young osteocytes or osteocytes entering apoptosis, but this percentage is prone to correspond to such a nature.

How many cilia-like structures per osteocyte ? In all our experiments, there was one and only one such a structure [12]. This is in accordance with the current knowledge on osteocytes [13, 14].

What size and ultrastructure for the centrosome? There were two centrioles: the mother centriole connected to the cell membrane, and daughter centriole never connected to the cell membrane. The size of the mother centriole average 482 ± 71 nm while the daughter centriole was smaller (351 ± 38 nm). The two centrioles were connected by striated rootlets in a large percent of cells, the mean distance between mother and daughter centrioles averaging 148 ± 74 nm.

What orientation for the centrosome?

In our experiment, the mother centriole connected to the cell membrane was oriented close to a parallel position to the section plane, which means an orientation perpendicular to the long axis of tibial bone.

This preferential orientation suggests a role in mechanosensing of this structure.

Morphology of the area connecting the mother centriole to the cell membrane

There was an electron-dense material in this area, with in some cells short cilium rods.

In one case, we observed a “cilium membrane rame” [12] between the cell body and the bone tissue. In cell-culture of osteocytes, such structures in extra cellular location have been described [15, 16, 17]. We have suggested that the specific bone environment could limit cilium growth [12]. The distal appendages are also located in this region, nine per mother centriole, one per triplet of the mother centriole.

We have also underlined [12] that the osteocyte differentiation was associated to a variation of the respective orientation of mother and daughter centriole.

The role of this ciliary structure in mechano-transduction has been underlined by several experiments: the loss of ciliary structure is characterized by the disappearance of prostaglandin expression in response to mechanical stress [16].

Another experiment has shown that the deletion of the PKD1 protein (part of the primary cilium structure) leads to a low bone mass, low anabolic processes, and bone defects [19]. Other studies confirm the evidence of these primary cilium structures' role in mechano-reception, conditioning bone formation and bone mass [19, 20]. Furthermore, the role of the ciliary structures in Wnt-signaling induced by mechanosensing has been demonstrated by hypogravity experiments

[21]. The cilia had previously been implicated in the regulation of Wnt-signaling [22, 23, 24].

In other organs, the primary cilium structures appear to be related to the calcium signaling [25, 26] while it does not seem to be the case in bone cells [27]. A direct role of the primary cilium structure on the PGE2 signaling has been suggested [27, 28, 29, 16].

In conclusion, the osteocyte primary cilium structure has been largely imaged by our recent study, and appears to play a major role in the mechano-sensing process in bone, considered as an early step in the bone formation process.

Acknowledgments The authors thank all the members of IPROS-I3MTO Laboratory for their help in this study.

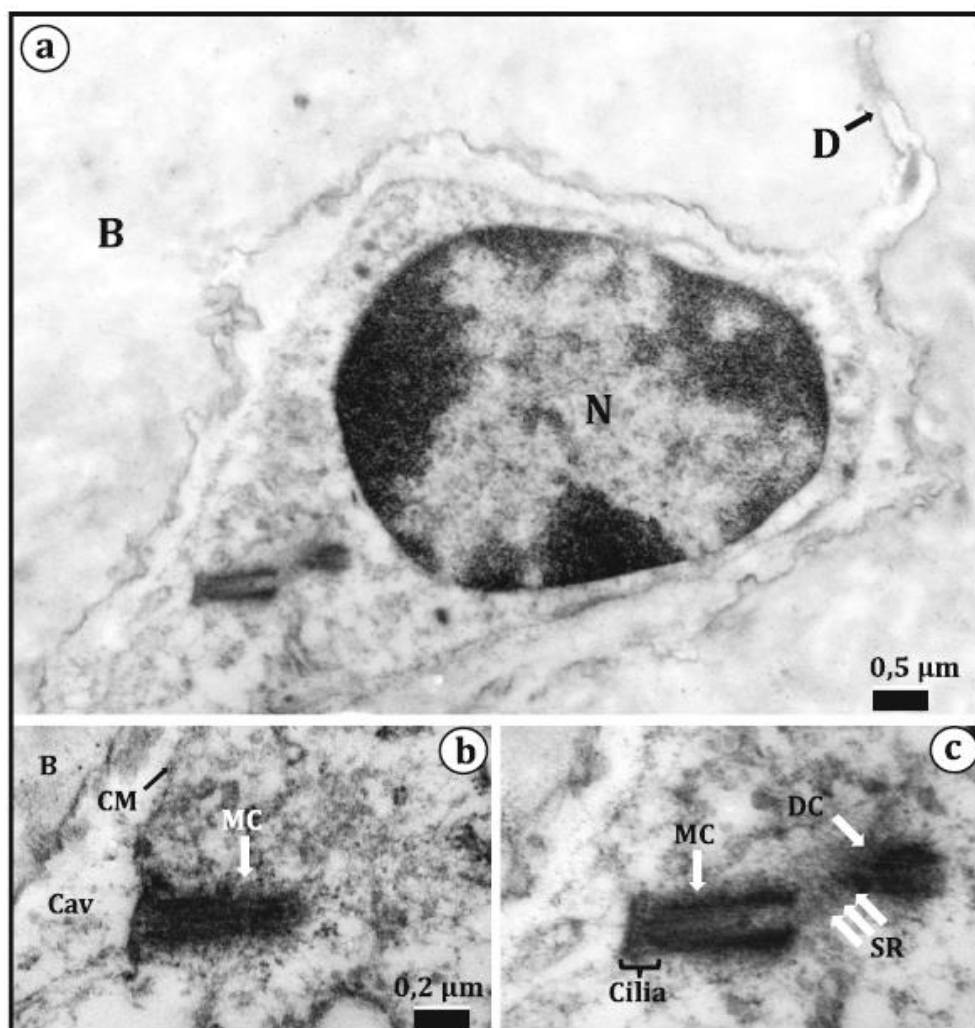


Fig. 1 The mother and daughter centriole (*MC*, *DC*) with the striated rootlet (*SR*), the extra-cellular cavity (*cav*), the dendrite (*D*), the nucleus (*N*) and the bone matrix (*B*) in an osteocyte from male Wistar rat imaged by transmission electron microscopy (TEM)

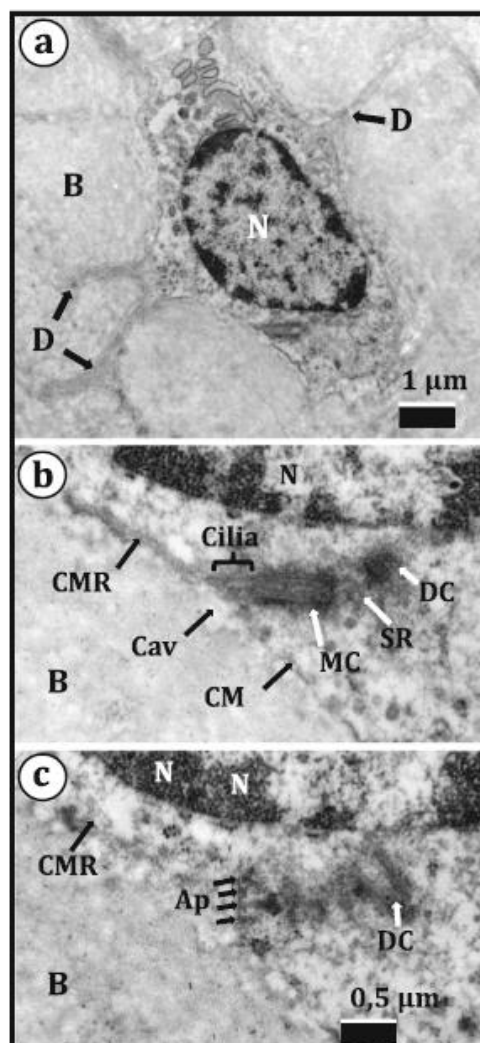


Fig. 2 Magnification of the junction between primary cilia and the extra cellular cavity with identification of the cilia membrane (CMR)

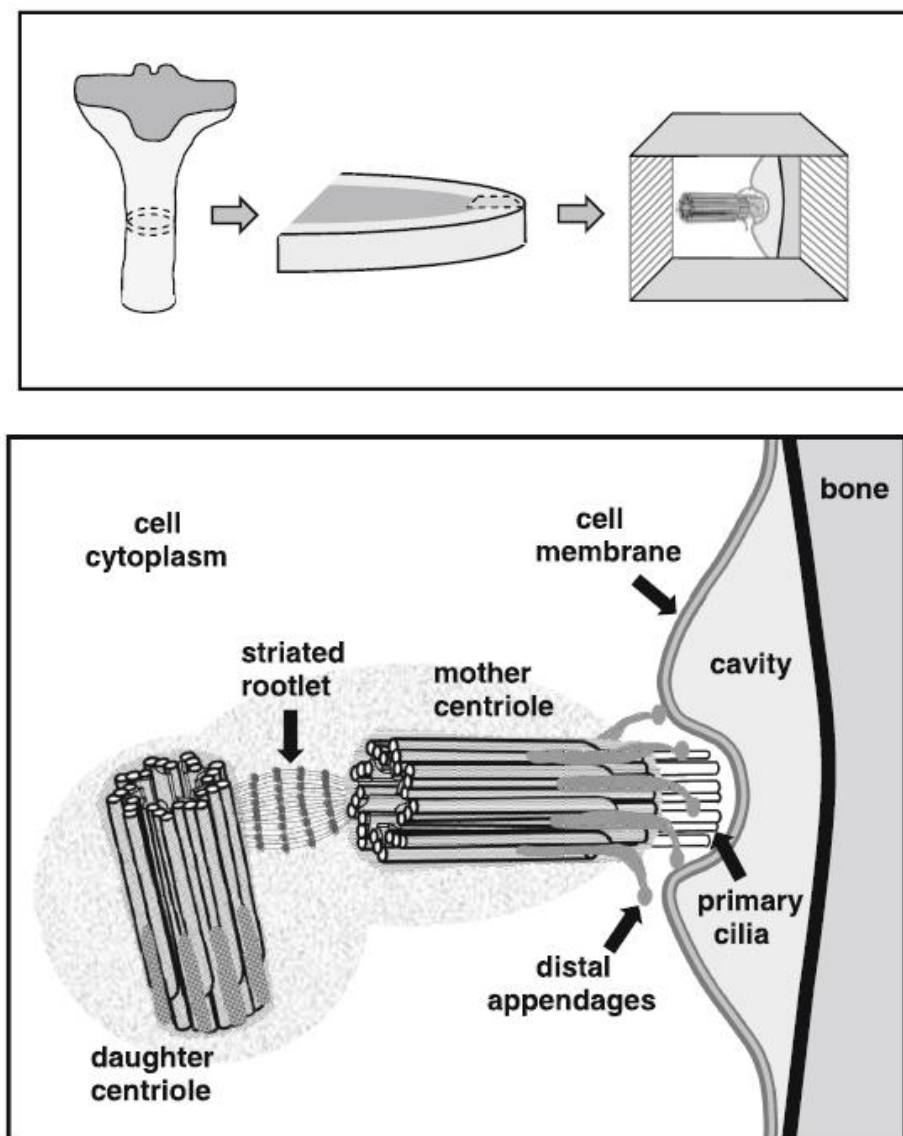


Fig. 3 Schematic representation of the centrosome with its fine microtubule constitution and its orientation relative to the rat tibia (adapted from 12)

Disclosure statement C.L. BENHAMOU has regularly worked with several pharmaceutical laboratories on different topics (clinical research, conferences, post-university teaching, boards) - Amgen- GSK- Lilly -Merck Sharp Dohme –Novartis-

Roche / Roche Chugai- Servier -UCB Pharma - Warner Chilcott – Wyeth – Rottapharm. The publication of the Eighth Bone Quality Seminar Proceedings 2013 has been made possible by an educational grant from Servier.

References

Osteocytes: an exquisitely sensitive mechanosensory cell

- Bonewald L. F. (2006) Mechanosensation and Transduction in Osteocytes. *Bonekey Osteovision* 3(10):7–15
- Bonewald L. F. (2011) The amazing osteocyte. *J Bone Miner Res* 26(2):229–238
- Bonewald LF, Johnson ML (2008) Osteocytes, mechanosensing and Wnt signaling. *Bone* 42(4):606–615
- Bonivitch AR, Bonewald LF et al (2007) Tissue strain amplification at the osteocyte lacuna: a microstructural finite element analysis. *J Biomech* 40(10):2199–2206
- Batra N, Burra S, Siller-Jackson AJ, Gu S, Xia X, Weber GF, DeSimone D, Bonewald LF, Lafer EM, Sprague E, Schwartz MA, Jiang JX (2012) Mechanical stress-activated integrin $\alpha 5 \beta 1$ induces opening of connexin 43 hemichannels. *Proc Natl Acad Sci USA* 109(9):3359–64.
- Burra S, Nicolella DP et al (2010) Dendritic processes of osteocytes are mechanotransducers that induce the opening of hemichannels. *Proc Natl Acad Sci USA* 107(31):13648–13653
- Cherian PP, Cheng B et al (2003) Effects of mechanical strain on the function of Gap junctions in osteocytes are mediated through the prostaglandin EP2 receptor. *J Biol Chem* 278(44):43146–43156
- Cherian PP, Siller-Jackson AJ et al (2005) Mechanical strain opens connexin 43 hemichannels in osteocytes: a novel mechanism for the release of prostaglandin. *Mol Biol Cell* 16(7):3100–3106
- Dallas SL, Prideaux M et al (2013) The Osteocyte: An Endocrine Cell and More. *Endocr Rev* 34:658–690
- Fritton SP, Weinbaum S (2009) Fluid and Solute Transport in Bone: Flow-Induced Mechanotransduction. *Annu Rev Fluid Mech* 41:347–374
- Genetos DC, Geist DJ et al. (2005) Fluid shear-induced ATP secretion mediates prostaglandin release in MC3T3-E1 osteoblasts. *J Bone Miner Res* 20(1):41–49
- Gu YM, Preston R et al (2001) Hormonally-regulated expression of voltage-operated Ca^{2+} channels in osteocytic (MLO-Y4) cells. *Biochem Biophys Res Commun* 282(2):536–542
- Javaheri B, Dallas M, Zhao H, Bonewald L, Johnson M (2011) b-Catenin Haploinsufficiency in Osteocytes Abolishes the Osteogenic Effect of Mechanical Loading in Vivo. *J. Bone Min Res*: S24
- Jing D, Lu XL et al (2013) Spatiotemporal properties of intracellular calcium signaling in osteocytic and osteoblastic cell networks under fluid flow. *Bone* 53(2):531–540
- Kalogeropoulos MS, Varanasi S et al (2010) Zic1 transcription factor in bone: neural developmental protein regulates mechanotransduction in osteocytes. *FASEB J* 24(8):2893–2903
- Kamel MA, Picconi JL et al (2010) "Activation of beta-catenin signaling in MLO-Y4 osteocytic cells versus 2T3 osteoblastic cells by fluid flow shear stress and PGE2: Implications for the study of mechanosensation in bone. *Bone* 47(5):872–881
- Kamioka HY, Miki et al. (1995). Extracellular calcium causes the release of calcium from intracellular stores in chick osteocytes. *Biochem Biophys Res Commun* 212(2):692–696
- Kitase Y, Barragan L et al (2010) Mechanical induction of PGE(2) in osteocytes blocks glucocorticoid induced apoptosis through both the beta-catenin and PKA pathways. *J Bone Miner Res* 25:2657–2668
- Klein-Nulend JA, Bakker D et al (2013) Mechanosensation and transduction in osteocytes. *Bone* 54(2):182–190
- Klein-Nulend J, Semeins et al (1995a) Pulsating fluid flow increases nitric oxide (NO) synthesis by osteocytes but not periosteal fibroblasts—correlation with prostaglandin upregulation. *Biochem Biophys Res Commun* 217(2):640–648
- Klein-Nulend J, van der Plas A et al (1995b) Sensitivity of osteocytes to biomechanical stress in vitro. *Faseb J* 9(5):441–445
- Kramer I, Halleux C et al (2010) Osteocyte Wnt/beta-catenin signaling is required for normal bone homeostasis. *Mol Cell Biol* 30(12):3071–3085
- Li J, Liu D et al (2005) The P2X7 nucleotide receptor mediates skeletal mechanotransduction. *J Biol Chem* 280(52):42952–42959
- Lin C, Jiang X et al (2009) Sclerostin mediates bone response to mechanical unloading through antagonizing Wnt/beta-catenin signaling. *J Bone Miner Res* 24(10):1651–1661
- Lu XL, Huo B et al (2011) Osteocytic network is more responsive in calcium signaling than osteoblastic network under fluid flow. *J Bone Miner Res*
- Malone AM, Anderson CT et al (2007) Primary cilia mediate mechanosensing in bone cells by a calcium-independent mechanism. *Proc Natl Acad Sci USA* 104(33):13325–13330
- Price C, Zhou X, Li W, Wang L (2010) Real-time measurement of solute transport within the lacunar-canalicular system of mechanically loaded bone: direct evidence for load-induced fluid flow. *J Bone Min Res* in press
- Robling AG, Niziolek PJ et al (2008) Mechanical stimulation of bone in vivo reduces osteocyte expression of Sost/sclerostin. *J Biol Chem* 283(9):5866–5875
- Sawakami K, Robling AG et al (2004) Site-specific osteopenia and decreased mechanoreactivity in Lrp5 mutant mice. *J Bone Min Res* 19(S1):S38
- Thompson WR, Majid AS et al (2011) Association of the $\alpha 2 \delta 1$ subunit with $\text{Ca}_v 3.2$ enhances membrane expression and regulates mechanically induced ATP release in MLO-Y4 osteocytes. *J Bone Miner Res* 26(9):2125–2139
- Uzbekov RE, Maurel DB, Aveline PC, Pallu S, Benhamou CL, Rochefort GY (2013) Centrosome fine ultrastructure

- of the osteocyte mechanosensitive primary cilium. *Microsc Microanal* in press:1–12
32. Weinbaum S, Cowin SC et al (1994) A model for the excitation of osteocytes by mechanical loading-induced bone fluid shear stresses. *J Biomech* 27(3):339–360
 33. Weinbaum S, Duan Y, Thi MM, You L (2011) An Integrative Review of Mechanotransduction in Endothelial, Epithelial (Renal) and Dendritic Cells (Osteocytes). *Cellular and Molecular Bioengineering* 4(4):510–537
 34. Xia X, Batra N et al (2010) Prostaglandin promotion of osteocyte gap junction function through transcriptional regulation of connexin 43 by glycogen synthase kinase 3/beta-catenin signaling. *Mol Cell Biol* 30(1):206–219
 35. Xiao Z, Dallas M et al (2011) Conditional deletion of Pkd1 in osteocytes disrupts skeletal mechanosensing in mice. *FASEB J* 25(7):2418–2432
 36. Xiong J, Onal M et al (2011) Matrix-embedded cells control osteoclast formation. *Nat Med* 17(10):1235–1241

Received: 25 June 2014 / Accepted: 25 July 2014

© Springer 2014

Reprints: Springer

Springer

Springer

Springer

Springer

Springer

Springer

Springer

Springer

Springer

Springer

Springer

Springer

Springer

Springer

Springer

Springer

Springer

Springer

Springer

Springer

Springer

Springer

Springer

Springer

Springer

Springer

Springer

Springer

Springer

Springer

Springer